

Selected Problems of Malaria Blood Stage Immunity

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Summary

Both antibody dependent and cell mediated mechanisms contribute to immunity in malaria. The parasites vary in sensitivity to antibody mediated inhibition due to underlying antigenic variation. When *Plasmodium falciparum* isolates are tested with antibodies from the donor originally harbouring the parasites or with those from other donors, growth inhibition is usually lowest in the autologous combinations. Parasites with decreased sensitivity are also generated *in vitro* by culturing them for prolonged periods in the presence of certain anti-plasmodial antibodies. When the antibodies are removed, a successive return of sensitivity develops. The decrease in sensitivity to inhibition may either be due to down-regulation of synthesis of the antigen or a selection of parasites with low antigen expression from the heterogeneous original populations.

Both T lymphocytes carrying α/β and γ/δ antigen-receptors play a role in malaria immunity. However, although γ/δ T cells may expand 40-fold or more in the peripheral immune system in acutely infected humans and also inhibit parasite growth *in vitro* and *in vivo*, their relative importance for protection or pathogenicity is presently unclear. Of the two major T cell subsets ($CD4^+$, $CD8^+$) carrying α/β T cell receptors, the role of $CD8^+$ T cells in blood stage infections appears to be limited. Instead, $CD4^+$ T cells are of major importance. These cells comprise at least two functionally different subsets (Th1, Th2), distinguished on the basis of lymphokine secretion. In some rodent malaria models, Th1 cells producing primarily IL2 and IFN γ give rise to protection in early infection while Th2 cells producing IL4 are essential for parasite clear-

ance in late infection. In other mouse strains, the same parasites induce a strong Th2 response in early infection, resulting in a lethal course. $CD4^+$ T cells of either Th1 or Th2 type also have regulatory functions in human *P. falciparum* malaria.

Most humans living in areas of high endemicity have significantly elevated blood levels of IgE, reflecting a skewing of the underlying T helper cell ratio in favour of Th2, responsible for the switch in immunoglobulin isotypes. Less than 5% of the IgE in malaria represents antibodies to *P. falciparum*. IgE elevation is highest in patients with severe and particularly cerebral malaria and is frequently associated with an elevation of tumour necrosis factor alpha (TNF). The release of this cytokine from monocytes/macrophages may reflect cross-linking of their low affinity receptors for IgE (CD23) by IgE containing immune complexes from malarial sera. Local overproduction of TNF is considered a major pathogenic mechanism, responsible for fever and tissue lesions in severe malaria. Although TNF overproduction in malaria is generally assumed to be due to direct stimulation of effector cells by certain parasite derived toxins, the present results suggest that IgE elevation constitutes yet another mechanism contributing to the pathogenicity of *P. falciparum* in human malaria.

Introduction

Malaria infection gives rise to immune responses with the ultimate goal to free the host from the infecting parasites. However, immune responses may also elicit reactions that are harmful to the host and the balance between protective and pathogenic responses will greatly affect the final outcome of an infection. In this brief survey we will discuss

selected aspects of the immune mechanisms regulating protection or pathogenicity in malaria blood stage infection, that is the phase responsible for mortality and morbidity of the disease.

Blood stage immunity and host genetics

Immunity to malaria infection is stage- and species specific. Acquisition of efficient blood stage immunity is relatively slow, age dependent and immunity is not lasting, i. e. acquired protection may be lost upon cessation of exposure (1). The reasons for this are multiple but the genetic diversity and variability of potentially protective parasite antigens is thought to be of major importance (2, 3). The development of immunity is also dependent on the genetics of the human host. Thus, the importance of particular HLA alleles for resistance to severe malaria has been reported several years ago (4). However, other genetic factors are also involved (5, 6). Of particular interest are the recent findings by Modiano *et al* (7) who studied *P. falciparum* malaria in populations belonging to three ethnic groups but living under similar conditions of hyperendemic malaria exposure in Burkina Faso. Of these groups, individuals belonging to one of them (Fulani) were apparently less susceptible to *P. falciparum* malaria as reflected by both lower parasitemias and fewer clinical episodes. Furthermore, this partial resistance was associated with an increased capacity to form antibodies against the parasite (8). As these enhanced humoral immune responses were polyspecific and directed against many parasite antigens, it is unlikely that the heightened reactivity reflected a genetic regulation by the MHC system. Further elucidation of the molecular-genetic basis of this improved responsiveness (which may not be unique for plasmodial infections) will be of great interest.

Antibodies and antigen expression

Although antibodies have long been known to protect against human *P. falciparum* malaria, the relative importance of different antibodies requires further explorations (for review and references see (9)). Moreover, by exhibiting antigenic diversity and variation, the parasites have developed mechanisms to escape the immune response (10). This may be illustrated by recent stud-

ies by Wåhlin *et al.* (11) who investigated merozoite invasion/growth inhibition *in vitro* of antibodies from children living in a *P. falciparum* malaria hyperendemic area of Burkina Faso. When comparing inhibitory antibody activities on autologous or heterologous parasites, a significantly lower inhibition was seen in autologous combinations. Principally similar results were recently obtained by J. Iqbal *et al.* (12) who cultured a *P. falciparum* laboratory strain (F32) *in vitro* under prolonged periods in the presence of suboptimal concentrations of a human monoclonal antibody against the blood stage antigen Pf332 (13, 14). In freshly isolated parasites, this antibody inhibited growth but prolonged culturing resulted in relative resistance to inhibition. No change in growth inhibition was seen when parasites cultured with this monoclonal antibody were tested for growth inhibition by antibodies specific for other antigens. Continued culturing of the parasites in the absence of antibody indicated that resistance to inhibition was reversible (12). Both examples illustrate how the parasites may defend themselves against the host's immune response. Whether this defense is based on direct down-regulation of antigen by antibody pressure or reflects selection of parasites with low antigen expression out of a heterogeneous population is presently unclear.

There are many pathways for antibody mediated protection in malaria, with opsonization of infected erythrocytes and ensuing phagocytosis or cytotoxicity by various effector cells probably being among those of greatest importance (15). There is presently no evidence that invasion/ growth inhibition by antibodies without effector cell involvement plays any major role in protection. However, the capacity of the parasites to change their antigenic set-up under pressure as described above is obviously of general validity regardless of which antibody mediated mechanisms may be involved in protection.

T-cell regulation and cell mediated immunity in malaria

γ/δ T cells. Normally, T cells equipped with γ/δ T cell receptor (TCR) comprise 5-10% of the circulating T cell pool. However, in acute infection this percentage may rise

up to 40%, reflecting not only a relative but also an absolute increase in numbers (16-18). This elevation which also has been reported for *P. vivax* infection (19) may persist for many weeks. Similar γ/δ T cell elevations have also been seen in other infections but the function of these cells is still a matter of speculation. Frequently, activation of γ/δ T cells leads to the release of Th1 cytokines (20) and an associated overproduction of IFN γ and TNF may become pathogenic. However, transfer of cloned γ/δ T cells from mice immunized with *P. yoelii* sporozoites to α/β TCR deficient mice has been shown to inhibit parasite development in the liver (21). There is no clear evidence suggesting that γ/δ T cells protect against blood stage infection. Nevertheless, γ/δ T cells from malaria naive humans activated by PHA or by culture with *P. falciparum* lysates also inhibit parasite growth in RBC *in vitro* in a cell contact requiring reaction (22). The crucial question of what these killer T cells see on the infected erythrocytes is unresolved. Considering what is known from other systems, it could be heat shock proteins exposed on the stressed (i. e. infected) erythrocytes or, rather, some phosphate containing glycolipids (23, 24). Findings to this effect have recently also been made in malaria (25, 26).

α/β T cells. The γ/δ T cell activities described above require no foregoing immunization and are not MHC-restricted. Thus, γ/δ T cells are considered to constitute a first line of defense which is rapidly mobilized. However, most known T cell activities in plasmodial infections involve T cells equipped with α/β receptors. These cells which normally are the majority of the peripheral T cells appear to have a larger repertoire than the γ/δ T cells and are MHC restricted in their reactivities with antigens. They have both cytotoxic and regulatory functions.

In malaria, T cell cytotoxicity by MHC class I restricted CD8⁺ T cells (27) and perhaps also by MHC class II restricted CD4⁺ T cells (28) appears to play a role in the defense against pre-erythrocytic infection in the liver. However, in the ensuing blood infection, cytotoxic T cells seem to have no role, in line with the poor expression of

MHC antigens on the erythrocytes, the main targets of the parasites. Thus, in the immune response during the erythrocytic phases of plasmodial infections, regulatory CD4⁺ T cells are the main players (29).

T cell polarization

In the mouse it has been known for several years that long lasting stimulation of CD4⁺ T cells results in functional polarization into at least two (but probably more) cell types, the so-called T helper 1 (Th1) and T helper 2 (Th2) cells. Later, it has become clear that similar distinctions are true for human CD4⁺ cells as well (30, 31) and can also be made for CD8⁺ T cells (32). Th1 and Th2 cells are distinct with regard to the cytokines they produce and, therefore, with regard to function (33). Upon activation, all T cells produce a large variety of cytokines. CD4⁺ Th1 cells that are involved in macrophage activation, cellular cytotoxicity and delayed type hypersensitivity, produce the key cytokines interferon gamma (IFN γ) and interleukin 2 (IL2). For Th2 cells which control humoral immunity and have major roles in allergy, asthma and certain infections, the key lymphokines are IL4, IL5 and IL13 (30, 34).

In "classical" studies performed by J. Langhorne and colleagues about 10 years ago (35) it was shown in a non-lethal mouse malaria system (*P. chabaudi chabaudi* AS) that the first immune response in naive mice against the parasite was a Th1 response, dominated by IFN γ and IL2 producing lymphocytes in the spleen. After 10-14 days when the infection had peaked, this response changed into a Th2 response dominated by IL4 and certain antibodies, finally resulting in complete clearance of the infection.

Host- and parasite factors affecting T cell responses

Several factors have been shown to affect the course of T cell responses and thereby the outcome of malaria infections. Studies similar to those referred to above but performed in mice differing in resistance of susceptibility to the same parasite (*P. chabaudi*) demonstrated the importance of host genetics (36). Thus, while in resistant mice the sequence of responses was as above (Th1 followed by Th2), in susceptible mice the

first response was Th2; such mice died of severe anemia and fulminant parasitemia. These results reflect the fact that protection against liver infection requires cell mediated reactions involving CD4⁺ T cells of Th1 type and cytotoxic CD8⁺ T cells.

Not only host- but also parasite factors will affect the development of the immune response. This may be exemplified by infection of mice with *P. vinckei* which is lethal and causes an illness involving overproduction of NO and TNF (37), i. e. a harmful Th1 response. When rescued by drug cure, they develop a solid immunity against reinfection, independently of antibodies or CD8⁺ cells (38). Rather, it is due to a protective Th1 response, characterized by splenocytes that upon antigen stimulation produce IFN γ but no IL4 (39). Why *P. vinckei* that closely resembles *P. chabaudi* only elicits Th1 responses while *P. chabaudi* infected mice appear to be rescued by the shift of a Th1 into a Th2 response is an important but unresolved issue.

IgE in human *P. falciparum* infection

More than 85% of individuals living in malaria endemic areas have elevated blood levels of IgE, roughly corresponding to the endemicity of the region (40, 41). Of this IgE less than 5% represents antibodies against many different plasmodial antigens. An IgE elevation appearing after repeated infections has also been found in the murine *P. chabaudi* model (42).

The formation of IgE and other immunoglobulin isotypes is controlled by T cells. Thus, the Ig isotype patterns found in serum may reflect various T helper cell reactivities (43). The switching of IgM (and IgG) to IgE is induced by the cytokines IL4 and IL13, released from activated Th2 cells (44). In line with this, *in vitro* activation of T cells from peripheral blood of malaria sensitized donors with antigen or mitogen (PHA) results in an increased ratio of IL4- over IFN γ -releasing cells, reflecting the presence of increased numbers of Th2 cells (45). The Th1 to Th2 switch is controlled by many factors including antigen dose and structure, length of antigen exposure, mode of immunization and host genetics (46). The latter is illustrated by studies of African twins, in

whom IgE levels were very similar in homozygous twin pairs but varied widely in heterozygous pairs (M. Troye-Blomberg *et al.*, manuscript in preparation). These results confirm previous more general results on antibody and T cell responses of malaria exposed homozygous or heterozygous twins (5, 6). The genes regulating these responses were located both within and outside the MHC region.

Role of IgE in malaria infection

In infectious disease IgE usually acts on parasites or parasitized cells through the intervention of effector cells. Interaction of IgE containing immune complexes with Fc ϵ receptors on different types of effector cells is known to induce a variety of responses which can be protective and/or harmful for the host (47). There are two major types of Fc ϵ receptors mediating IgE dependent cellular reactions. The so-called high affinity IgE receptor, Fc ϵ RI, occurs primarily (but not exclusively) on mast cells and basophils and is the major mediator of immediate hypersensitivity reactions (48). The other receptor, Fc ϵ RII or CD23, occurs primarily on monocytes/macrophages but also on B cells and other hematopoietic cells (49). This receptor, also called low affinity receptor, is induced to elevated expression by IgE and/or IL4 (50, 51) and may mediate IgE dependent phagocytosis, cellular cytotoxicity and adhesion (52-54). IgE dependent reactions encountered, for example, in helminthic infections but also in malaria, are believed to be primarily mediated by CD23.

While IgE's possible protective effects in malaria have not as yet been investigated, there is evidence for its involvement in pathogenesis. This is based on the initial findings in cerebral as well as in severe non-cerebral malaria of significantly higher IgE concentrations than in the blood of uncomplicated cases. This IgE elevation included both total and anti-parasitic IgE while no similar elevations were found for IgG (41, 55). With regard to specificity, the anti-plasmodial IgE antibodies in severe malaria cases appear to be qualitatively the same as in uncomplicated cases, indicating that the elevations seen in severe malaria were not due to IgE responses to some "disease specific" antigens.

IgE and TNF in malaria pathogenesis

Two not mutually exclusive reaction patterns are usually implicated in the pathogenesis of cerebral malaria: one involves obstruction of blood flow in the postcapillary microvasculature in the brain (56) due to sequestration of parasitized erythrocytes and resulting in ischaemia, acidosis, anoxia, ring hemorrhages and tissue necrosis. The other involves tissue damaging effects of cytokines, reactive nitrogen or oxygen, induced by the interaction of parasites with various leukocytes or vessel endothelium (57). One of the important cytokines released under such conditions is tumor necrosis factor α (TNF). Although TNF may protect against parasites (58-60), elevated concentrations are correlated with disease severity (61-63) and in cerebral malaria even with mortality (64). It is also a major factor inducing malaria fever (65).

TNF is released from monocytes, activated by, for example, bacterial lipopolysaccharides. In malaria, parasite pigment, various antigens and toxins released from the parasite have also been implicated in TNF induction (66, 67). Recently, major attention has been given to the GPI (=glycosylphosphatidyl-inositol) molecules that are anchoring many polypeptides, including malaria exoantigens, in the cell membrane (68-70). However, monocytes can be activated to produce cytokines and various effector functions via different pathways. Thus, as already indicated, the surface expression on monocytes/macrophages of Fc ϵ RII, CD23, is upregulated by IL4 and/or IgE (50, 51) and cross-linking of this receptor by IgE containing immune complexes results in the production of nitric oxide and TNF (53) which both are effector molecules in the defense against the parasite. In line with this, IgE and TNF elevation frequently parallel each other in serum of severe malaria patients (55). At the same time, sequestration of *P. falciparum* infected erythrocytes to small vessels of the brain in cerebral malaria will also result in the deposition of IgE containing immune complexes (Y. Maeno *et al.*, manuscript in preparation), assumedly leading to local overproduction of TNF and ensuing tissue lesions. It should be emphasized here that it is not the high levels of

TNF alone that causes the lesions typical for cerebral malaria. Other human malaria parasites such as *P. vivax* do not cause cerebral malaria but may induce much more TNF than *P. falciparum* (71). Rather, it would seem to be the combination of sequestration to capillaries and post-capillary venules of this parasite with local monocyte activation that is the basis for these serious complications of *P. falciparum* malaria.

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REFERENCES

- 1) Baird J. K., 1995. Host age as a determinant of naturally acquired immunity to *Plasmodium falciparum*. *Parasitol. Today* 11:105-111.
- 2) Anders R. F., 1986. Multiple cross-reactivities amongst antigens of *Plasmodium falciparum* impair the development of protective immunity against malaria with special reference to oxidant stress. *Parasite Immunol.* 8:529-539.
- 3) Walliker D., 1994. The role of molecular genetics in field studies on malaria parasites. *Int. J. Parasitol.* 24:799-808.
- 4) Hill A. V. S., Allsopp C. E. M., Kwiatkowski D., Anstey N. M., Twumasi P., Rowe P. A., Bennett S., Brewster D., McMichael A. J. and Greenwood B. M., 1991. Common West African HLA antigens are associated with protection from severe malaria. *Nature* 352:595-600.
- 5) Sjöberg K., Lepers J. P., Raharimalala L., Larsson Å., Olerup O., Marbian N. T., Troye-Blomberg M. and Perlmann P., 1992. Genetic regulation of human anti-malarial antibodies in twins. *Proc. Natl. Acad. Sci. USA* 89:2101-2104.
- 6) Jepson A., Banya W., Sisay-Joof F., Hassan-King M., Nunes C., Bennett S. and Whittle H., 1997. Quantification of the relative contribution of major histocompatibility complex (MHC) and non-MHC genes to human immune responses to foreign antigens. *Infect. Immunology* 65:872-876.
- 7) Modiano D., Petrarca V., Sirima B. S., Nebié I., Diallo D., Esposito F. and Coluzzi M., 1996. Different response to *Plasmodium falciparum* malaria in West African sympatric ethnic groups. *Proc. Natl. Acad. Sci. USA* 93:13206-13211.
- 8) Modiano D., Chiuchiuni A., Petrarca V., Sirima B. S., Luoni G., Perlmann H., Esposito F. and Coluzzi M.,

1997. Humoral response to *Plasmodium falciparum* PF155/RESA and Pf332 in three sympatric ethnic groups of Burkina Faso, West Africa. *Am. J. Trop. Med. Hyg.* in press.
- 9) Berzins K. and Perlmann P., 1996. Malaria vaccines: attacking infected erythrocytes. In Hoffman SL, ed. *Malaria Vaccine Development: A Multi-Immune Response Approach*. American Society for Microbiology Press, Washington, DC, 105-143.
 - 10) Reeder J. C. and Brown G. V., 1996. Antigenic variation and immune evasion in *Plasmodium falciparum* malaria. *Immunol. Cell Biol.* 74:546-554.
 - 11) Wählin Flyg B., Perlmann H., Perlmann P., Esposito F. and Berzins K., 1997. Wild isolates of *Plasmodium falciparum* malaria show decreased sensitivity to *in vitro* inhibition of parasite growth mediated by autologous host antibodies. *Clin. Exp. Immunol.* 107:321-327.
 - 12) Iqbal J., Siripoon N., Snounou G., Perlmann P. and Berzins K., 1997. *Plasmodium falciparum*: selection of parasite subpopulations with decreased sensitivity for antibody-mediated growth inhibition *in vitro*. *Parasitol.* 114:317-324.
 - 13) Mattei D. and Scherf A., 1992. The Pf332 gene of *Plasmodium falciparum* codes for a giant protein that is translocated from the parasite to the membrane of infected erythrocytes. *Gene* 110:71-79.
 - 14) Ahlborg N., Berzins K. and Perlmann P., 1991. Definition of the epitope recognized by the *Plasmodium falciparum*-reactive human monoclonal antibody 33G2. *Mol. Biochem. Parasitol.* 46:89-96.
 - 15) Bouharoun-Tayoun H., Attanath P., Sabchareon A., Chongsuphajaisiddhi T. and Druilhe P., 1990. Antibodies that protect humans against *Plasmodium falciparum* blood stages do not on their own inhibit parasite growth and invasion *in vitro*, but act in cooperation with monocytes. *J. Exp. Med.* 172:1633-1641.
 - 16) Ho M., Tongtawe P., Kriangkum J. K., Wimouwattra-Watee T., Pattanapanyasat K., Bryant L., Shafiq J., Suntharsamal S., Loareesuwan S., Webster H. K. and Eliot J. F., 1994. Polyclonal expansion of peripheral γ/δ + T cells in human *Plasmodium falciparum* malaria. *Infect. Immunity* 62:855-862.
 - 17) Goodier M., Krausejauer M., Sanni A., Massougbojji A., Sadeler B. C., Mitchell G. H., Modolell M., Eichmann K. and Langhorne J., 1993. γ/δ -T cells in the peripheral blood of individuals from an area of holoendemic *Plasmodium falciparum* transmission. *Trans. Roy. Soc. Trop. Med. Hyg.* 87:692-696.
 - 18) Roussilhon C., Agrapart M., Ballet J. J. and Bensussan A., 1990. T lymphocytes bearing the γ/δ T cell receptor in patients with acute *Plasmodium falciparum* malaria. *J. Infect. Dis.* 162:283-285.
 - 19) Perera M. K., Carter R., Goonewardene R. and Mendis K. M., 1994. Transient increase in circulating γ/δ T cells during *Plasmodium vivax* malarial paroxysms. *J. Exp. Med.* 179:311-315.
 - 20) Goodier M. R., Lundqvist C., Hammarström M.-L., Troye-Blomberg M. and Langhorne J., 1995. Cytokine profiles for human V γ 9⁺ T cells stimulated by *Plasmodium falciparum*. *Parasite Immunol.* 17:413-423.
 - 21) Tsuji M., Mombaerts P., Lefrancois L., Nussenzweig R. S., Zavala F. and Tonegawa S., 1994. γ/δ T cells contribute to immunity against the liver stages of malaria in $\alpha\beta$ T-cell-deficient mice. *Proc. Natl. Acad. Sci. USA* 91:345-349.
 - 22) Elloso M. M., van der Heyde H. C., vande Waa J. A., Manning D. D. and Weidanz W. P., 1994. Inhibition of *Plasmodium falciparum* *in vitro* by human γ/δ T cells. *J. Immunol.* 153:1187-1194.
 - 23) Kaufmann S. H. E., 1996. γ/δ and other unconventional T lymphocytes: what do they see and what do they do? *Proc. Natl. Acad. Sci. USA* 93:2272-2279.
 - 24) Tanaka Y., Morita C. T., Tanaka Y., Nicoes E., Brenner M. B. and Bloom B. R., 1995. Natural and synthetic non-peptide antigens recognized by human γ/δ T cells. *Nature* 375:155-158.
 - 25) Behr C., Poupot R., Peyrat M.-A., Poquet Y., Constant P., Dubois P., Bonneville M. and Fournie J.-J., 1996. *Plasmodium falciparum* stimuli for human γ/δ T cells are related to phosphorylated antigens of mycobacteria. *Infect. Immun.* 64:2892-2896.
 - 26) Pichyangkul S., Saengkrai P., Yongvanitchit K., Stewart A. and Heppner D. G., 1997. Activation of γ/δ T cells in malaria: interaction of cytokines and a schizont-associated *Plasmodium falciparum* antigen. *J. Infect. Dis.* 176:233-241.
 - 27) Hoffman S. L., Franke E. D., Hollingdale M. R. and Druilhe P., 1996. Attacking infected hepatocytes. In Hoffman, S. L., ed., *Malaria Vaccine Development: A multi-immune response approach*. American Society for Microbiology Press, Washington, DC, 35-75.
 - 28) Moreno A., Clavijo P., Edelman R., Davis J., Szein M., Sinigaglia F. and Nardin E., 1993. CD4⁺ T-cell clones obtained from *Plasmodium falciparum* sporozoite-immunized volunteers recognize polymorphic sequences of the circumsporozoite protein. *J. Immunol.* 151:489-499.
 - 29) Troye-Blomberg M. and Perlmann P., 1994. Malaria immunity: an overview with emphasis on T cell function. Good MF and Saul AJ, eds. *Molecular Immunological Considerations in Malaria Vaccine Development*. Boca Raton, Florida, USA: CRC Press, Inc., 1-46.
 - 30) Mosmann T. R. and Coffman R. L., 1989. TH1 and TH2 cells: Different patterns of lymphokine secretion lead to different functional properties. *Annu. Rev. Immunol.* 7:145-173.
 - 31) Del Prete G. F., De Carli M., Mastromauro C., Biagiotti R., Macchia D., Falagiani P., Ricci M. and Romagnani S., 1991. Purified protein derivative of *Mycobacterium tuberculosis* and excretory-secretory antigen(s) of *Toxocara canis* expand *in vitro* human T cells with stable and opposite (Type 1 T helper or Type 2 T helper) profile of cytokine production. *J. Clin. Invest.* 88:346-350.
 - 32) Sad S., Marcotte E. and Mosmann T. R., 1995. Cytokine induced differentiation of precursor mouse CD8⁺ T cells into cytotoxic CD8⁺ T cells secreting Th1 or Th2 cytokines. *Immunity* 2:271-279.
 - 33) Abbas A. K., Murphy K. M. and Sher A., 1996. Functional diversity of helper T lymphocytes. *Nature* 378:787-793.

- 34) Romagnani S., 1994. Lymphokine production by human T cells in disease states. *Annu. Rev. Immunol.* 12:227-257.
- 35) Langhorne J., Gillard S., Simon B., Slade S. and Eichmann K., 1989. Frequencies of CD4⁺ T cells reactive with *Plasmodium Chabaudi chabaudi*: distinct response kinetics for cells with Th1 and Th2 characteristics during infection. *Int. Immunol.* 1:416-424.
- 36) Stevenson M. M. and Tam M. F., 1993. Differential induction of helper T cell subsets during blood-stage *Plasmodium chabaudi* as infection in resistant and susceptible mice. *Clin. Exp. Immunol.* 92:77-83.
- 37) Clark I. A., MacMicking J. D., Gray K. M., Rockett K. A. and Cowden W. B., 1992. Malaria mimicry with tumor necrosis factor. Contrasts between species of murine malaria and *Plasmodium falciparum*. *Am. J. Pathol.* 140:325-336.
- 38) Kumar S., Good M. F., Dontfraid F., Vinetz J. M. and Miller L. H., 1989. Interdependence of CD4⁺ T cells and malarial spleen in immunity to *Plasmodium vinckei vinckei*. Relevance to vaccine development. *J. Immunol.* 143:2017-2023.
- 39) Perlmann H., Kumar S., Vinetz J. M., Kullberg M., Miller L. H. and Perlmann P., 1995. Cellular mechanisms in the immune response to malaria in *Plasmodium vinckei*-infected mice. *Infect. Immun.* 63:3987-3993.
- 40) Desowitz R. S., 1989. *Plasmodium*-specific immunoglobulin E in sera from an area of holoendemic malaria. *Trans. Roy. Soc. Trop. Med. Hyg.* 83:478-479.
- 41) Perlmann H., Helmbly H., Hagstedt M., Carlson J., Larsson P. H., Troye-Blomberg M. and Perlmann P., 1994. IgE elevation and IgE anti-malarial antibodies in *Plasmodium falciparum* malaria: association of high IgE levels with cerebral malaria. *Clin. Exp. Immunol.* 97:284-292.
- 42) Helmbly H., Perlmann H., Troye-Blomberg M. and Perlmann P., 1996. Immunoglobulin E elevation in *Plasmodium chabaudi* malaria. *Infect. Immun.* 64:1432-1433.
- 43) Finkelmann F. D., Holmes J., Katona I. M., Urban Jr. J. F., Beckmann M. P., Park L. S., Schooley K. A., Coffman R. L., Mosmann T. R. and Paul W. E., 1990. Lymphokine control of in vivo immunoglobulin isotype selection. *Annu. Rev. Immunol.* 8:303-333.
- 44) Zurawski G. and de Vries J. E., 1994. Interleukin 13, an interleukin 4-like cytokine that acts on monocytes and B cells but not on T cells. *Immunol. Today* 15:19-26.
- 45) ElGhazali G., Perlmann H., Rutta A. S. M., Perlmann P. and Troye-Blomberg M., 1997. Elevated plasma levels of IgE in *Plasmodium falciparum*-primed individuals reflect an increased ratio of IL-4 to interferon-gamma (IFN- γ)-producing cells. *Clin. Exp. Immunol.* 109:84-89.
- 46) Constant S. L. and Bottomly K., 1997. Induction of Th1 and Th2 CD4⁺ T cell responses: the alternative approaches. *Annu. Rev. Immunol.* 15:297-322.
- 47) Perlmann P., Perlmann H. and Troye-Blomberg M., 1996. IgE and TNF in malaria infection. Protection and pathogenicity: two sides of the same coin. *The Immunologist* 4/5:179-184.
- 48) Bonnefoy J. Y., Aubry J. P., Gauchat J. F., Graber P., Life P., Flores-Romo L. and Mazzei G., 1993. Receptors for IgE. *Curr. Op. Immunol.* 5:944-949.
- 49) Delespesse G., Suter U., Mossalayi D., Bettler B., Sarfati M., Hofstetter H., Kilcherr E., Debré P. and Dalloul A., 1991. Expression, structure and function of the CD23 antigen. *Adv. Immunol.* 49:149-191.
- 50) Vercelli D., Jabara H. H., Lee B. W., Woodland N., Geha R. S. and Leung D. Y., 1988. Human recombinant interleukin 4 induces Fc ϵ R2/CD23 on normal human monocytes. *J. Exp. Med.* 167:1406-1416.
- 51) Kawabe T., Takami M., Hosada M., Maeda Y., Sato S., Mayumi H., Mikawa M., Arai K. I. and Ydoi J., 1988. Regulation of Fc ϵ R2/CD23 gene expression by cytokines and specific ligands (IgE and anti Fc ϵ R2/CD23 monoclonal antibody). Variable regulation depending on cell types. *J. Immunol.* 141:1376-1382.
- 52) Voldoukis I., Issaly F., Fourcade C., Paul-Eugène N., Arock M., Kolb J. P., Alves da Silva O., Monjour L., Pouisot H., Tselentis Y., Dugas B., Debré P. and Mossalayi M. D., 1994. CD23 and IgE expression during the human immune response to cutaneous leishmaniasis: possible role in monocyte activation. *Res. Immunol.* 145:17-27.
- 53) Dugas B., Mossalayi D., Damais C. and Kolb J. P., 1995. Nitric oxide production by human monocytes: evidence for a role of CD23. *Immunol. Today* 16:574-580.
- 54) Lecoanet-Henchoz S., Gauchat J. F., Aubry J. P., Graber P., Life P., Paul-Eugène N., Ferrua B., Corbi A. L., Dugas B., Plater-Zyberk C. and Bonnefoy J. Y., 1995. CD23 regulates monocyte activation through a novel interaction with the adhesion molecules CD11b-CD18 and CD11c-CD18. *Immunity* 3:119-125.
- 55) Perlmann P., Perlmann H., Flyg Wählin B., Hagstedt M., ElGhazali G., Worku S., Fernandez V., Rutta A. S. M. and Troye-Blomberg M., 1997. Immunoglobulin E, a pathogenic factor in *Plasmodium falciparum* malaria. *Infect. Immun.* 65:116-121.
- 56) Berendt A. R., Turner G. D. H. and Newbold C. I., 1994. Cerebral malaria: the sequestration hypothesis. *Parasitol. Today* 10:412-414.
- 57) Clark I. A. and Rockett K. A., 1994. The cytokine theory of human cerebral malaria. *Parasitol. Today* 10:410-412.
- 58) Nussler A., Drapier J. C., Renia L., Pied S., Miltgen F., Gentilini M. and Mazier D., 1991. L-Arginine-dependent destruction of intrahepatic malaria parasites in response to tumor necrosis factor and/or interleukin-6 stimulation. *Eur. J. Immunol.* 21:227-230.
- 59) Kowanko I. C., Ferrante A., Clemente G. and Kumaratilake L. M., 1996. Tumor necrosis factor primes neutrophils to kill *Staphylococcus aureus* by an oxygen-dependent mechanism and *Plasmodium falciparum* by an oxygen-independent mechanism. *Infect. Immun.* 64:3435-3437.
- 60) Jacobs P., Radzich D. and Stevenson M. M., 1996. A Th1-associated increase in tumor necrosis factor alpha expression in the spleen correlates with resistance to blood-stage malaria in mice. *Infect. Immun.* 65:535-541.

- 61) Grau G. E., Taylor T. E., Molyneux M. E., Virima J. J., Vassalli P., Hommel M. and Lambert P.-H., 1989. Tumor necrosis factor and disease severity in children with *Plasmodium falciparum* malaria. *New Engl. J. Med.* 320:1586-1591.
- 62) Shaffer N., Grau G. E., Hedberg K., Davachi F., Lyamba B., Hightower A. W., Grennan J. G. and Nguyen-Dinh P., 1991. Tumor necrosis factor and severe malaria. *J. Inf. Dis.* 163:96-101.
- 63) McGuire W., Hill A. V., Allsopp C. E. M., Greenwood B. M. and Kwiatkowski D., 1994. Variation in the TNF- α promoter region associated with susceptibility to cerebral malaria. *Nature* 371:508-511.
- 64) Kwiatkowski D., Hill A. V. S., Sambou I., Twumasi P., Castracane J., Manogue K. R., Cerami A., Brewster D. R. and Greenwood B. M., 1990. TNF concentrations in fatal cerebral, non-fatal cerebral, and uncomplicated *Plasmodium falciparum* malaria. *Lancet* 336:1201-1204.
- 65) Kwiatkowski D., Molyneux M. E., Stephens S., Curtis N., Klein N., Pointaire P., Smit M., Allan R., Brewster D. R., Grau G. E. and Greenwood B. M., 1993. Anti-TNF therapy inhibits fever in cerebral malaria. *Q. J. Med.* 86:91-98.
- 66) Pichyangkul S., Saengkrai P. and Webster H. K., 1994. *Plasmodium falciparum* pigment induces monocytes to release high levels of tumor necrosis factor- α and interleukin-1 β . *Am. J. Trop. Med. Hyg.* 51:430-435.
- 67) Kwiatkowski D., 1995. Malarial toxins and the regulation of parasite density. *Parasitol. Today* 11:206-212.
- 68) Bate C. A. W., Taverne J. and Playfair J. H. L., 1992. Detoxified exoantigens and phosphatidylinositol derivatives inhibit tumor necrosis factor induction by malarial exoantigens. *Infect. Immun.* 60:1894-1901.
- 69) Schofield L., Vivas L., Hackett F., Gerold P., Schwarz R. T. and Tachado S., 1993. Neutralizing monoclonal antibodies to glycosylphosphatidylinositol, the dominant TNF- α -inducing toxin of *Plasmodium falciparum*: prospects for the immunotherapy of severe malaria. *Ann. Trop. Med. Parasitol.* 87:617-626.
- 70) Schofield L. and Tachado S. D., 1996. Regulation of host cell function by glycosylphosphatidylinositols of the parasitic protozoa. *Immunol. Cell Biol.* 74:555-563.
- 71) Karunaweera N. D., Grau G. E., Gamage P., Carter R. and Mendis K. N., 1992. Dynamics of fever and serum levels of tumor necrosis factor are closely associated during clinical paroxysms in *Plasmodium vivax* malaria. *Proc. Natl. Acad. Sci. USA* 89:3200-3203.